

A single macrolichen constitutes hundreds of unrecognized species

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Edited by Peter H. Raven, Missouri Botanical Garden, St. Louis, MO, and approved May 20, 2014 (received for review February 25, 2014)

The number of Fungi is estimated at between 1.5 and 3 million. Lichenized species are thought to make up a comparatively small portion of this figure, with unrecognized species richness hidden among little-studied, tropical microlichens. Recent findings, however, suggest that some macrolichens contain a large number of unrecognized taxa, increasing known species richness by an order of magnitude or more. Here we report the existence of at least 126 species in what until recently was believed to be a single taxon: the basidiolichen fungus *Dictyonema glabratum*, also known as *Cora pavonia*. Notably, these species are not cryptic but morphologically distinct. A predictive model suggests an even larger number, with more than 400 species. These results call into question species concepts in presumably well-known macrolichens and demonstrate the need for accurately documenting such species richness, given the importance of these lichens in endangered ecosystems such as paramos and the alarming potential for species losses throughout the tropics.

diversification | global diversity prediction | Hygrophoraceae

Fungi make up the second largest kingdom, with an estimated number of 1.5–3 million species (1–3). Lichenization plays an important role in fungal evolution, particularly in the Ascomycota, where lichens make up 30% of the currently recognized species (4–6). Transition toward a lichenized lifestyle appears to have taken place at least 10 times in the Ascomycota and 5 times in the Basidiomycota (7–9), but the distribution of lichen formers favors the Ascomycota, with the Basidiomycota accounting for less than 0.3% of all lichenized Fungi (7, 10). Altogether, ~18,000 lichenized species are currently accepted, but estimates suggest that this represents only 50–65% of the true species richness (4, 6).

Global species richness of lichenized Basidiomycota appears to be especially underestimated. The *Dictyonema* clade, which includes some of the best-known basidiolichens, until recently was considered to represent five species in a single genus, *Dictyonema* (11, 12). Subsequent taxonomic and molecular phylogenetic studies suggested that this concept did not reflect the true diversity in this clade (7, 12, 13). Currently, a total of 43 species are recognized in five genera (14, 15). Two genera, *Cora* and *Corella*, are foliose macrolichens, with a total of 16 species, corresponding to what was considered a single species, *Dictyonema glabratum* (11, 12, 16). This name is well known in the scientific community and even among nonspecialists and is included in the *Listing of Interesting Plants of the World* (17). The 16-fold increase in the number of species now recognized is a striking figure that even surpasses recent findings reported from the large macrolichens *Lobariella* and *Stictia* in the Ascomycota (18, 19). The dramatic change in the taxonomic concept of these basidiolichens has important implications for recognizing their role in ecosystem function and as model organisms. Species of *Cora* abound in tropical montane regions and, with their cyanobacterial photobionts capable of fixing atmospheric nitrogen, serve as biological fertilizers (20). *Cora* is also one of the best studied lichens in terms of ecomorphology, ecophysiology, and biochemistry (10, 21–28).

Sixteen Species: The Tip of the Iceberg?

The increase from 1 to 16 currently recognized species in *Cora* and *Corella* still probably does not adequately encompass the true species richness in this group, given the limited sequence data available until recently (7, 13). For the present study, we assembled a much larger data set of the internal transcribed spacer (ITS) barcoding locus for these two genera, with a total of 376 sequences, which is more than an order of magnitude larger than the most recent phylogenetic study (13). ITS provides a surprising amount of resolution and support in this group, and our analyses, based on topology, support, and correlation with phenotype features, suggest that the total number of species in this dataset is 126, a nearly eightfold increase over the currently recognized 16 species (Fig. 1 and Fig. S1). Single-gene species recognition using a coalescent approach suggests even more species to be present. Single- and multiple-threshold Generalized Mixed Yule Coalescent (GMYC) analysis using the *R splits* package (29) resulted in 73–84 multisequence and 71–86 singleton species, for a total of 144–170 species recognized in the dataset.

Notably, most of these are not cryptic species recognizable from molecular data only, but morphologically distinct taxa supported by phenotype features, substrate ecology, habitat preferences, and geographical distribution. Many are so distinctive that it is surprising that they have not been recognized before (Fig. 2).

Significance

Macrolichens are considered to be well known, including the tropical montane basidiolichen fungus *Dictyonema glabratum*, also known as *Cora pavonia*, an important component of threatened paramo ecosystems, where it acts as a biological fertilizer due to its ability to fix atmospheric nitrogen. This lichen was long believed to represent a single species, but after revising this number to 16 in two genera (*Cora* and *Corella*), here we show that at least 126 phylogenetically and morphologically distinct species are contained within this group, with statistical analysis predicting more than 400. Our findings underline the importance of accurately documenting species richness for conservation purposes and support the notion of neotropical paramos as hotspots of recent diversification in plants, animals, and fungi.

Author contributions: R.L., M.D.-F., F.B., B.M., and J.D.L. designed research; R.L., M.D.-F., M.S., P.M.G., F.B., B.M., A.Y.-A., J.L.C., L.F.C., and J.D.L. performed research; R.L. and P.M.G. contributed new reagents/analytic tools; R.L., M.D.-F., M.S., P.M.G., F.B., B.M., A.Y.-A., J.L.C., L.F.C., and J.D.L. analyzed data; and R.L., M.D.-F., F.B., and J.D.L. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

Data deposition: The sequences reported in this paper have been deposited in the GenBank database (see Table S1 for accession nos.). Sequence alignment and tree file are available from TreeBASE, <http://treebase.org> (Study ID no. 15783).

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This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1403517111/-DCSupplemental.

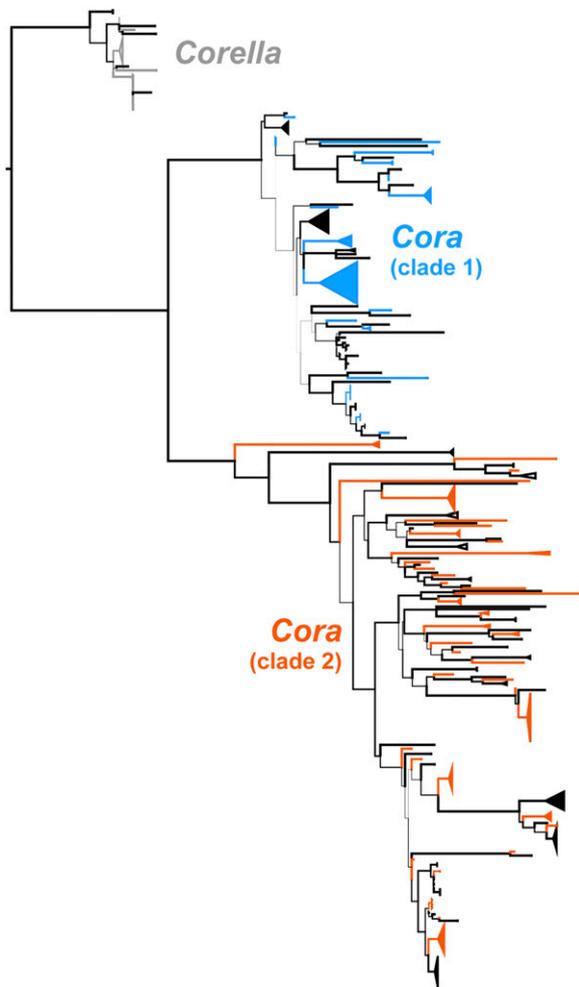


Fig. 1. Condensed, best-scoring maximum likelihood ITS phylogeny of *Cora* and *Corella*. Recognized species are indicated by alternating colors (blue-black for *Cora*, orange-black for *Corella*). See Fig. S1 for complete tree with bootstrap support values.

The likely explanation is that important features such as color, texture, and substrate are lost in dried herbarium specimens, which makes these lichens a prime example of taxa to be studied in the field. In instances where some of this diversity was acknowledged, it was attributed to infraspecific, habitat-induced variation of a single species (10, 24, 30), but our data show that this is not the case, and the correct interpretation of this variation has dramatic consequences for estimating taxonomic and phylogenetic diversity in this lineage.

Using a grid map of southern North America to South America that covers almost the entire distribution range of *Cora* and *Corella* (Fig. 3), we found that of the 126 recognized species, 101 were sampled within a single grid only. Another 16 taxa were sampled in two to five grids, and only one in more than five grids, suggesting a strong geographic signal and an unexpectedly high level of endemism in a lineage thought to represent a single, widely distributed species. This result does not appear to be a sampling artifact, because the number of sampled specimens was up to 18 per taxon in species found in one grid only and up to 17 per taxon in those found in more than one grid. Rather, it matches frequency patterns found in many other chiefly tropical organisms, with few frequent and many rare and often locally endemic species (31).

In addition, the recognition of such a large number of species does not appear to be an artifact of possible gene duplication, as

has been suggested for certain fungal lineages (32–36). In a separate analysis of 454 pyrosequencing data obtained from *Cora*, we found no evidence for ITS paralogs and instead demonstrated that almost all intragenomic variation can be attributed to sequencing errors (37). This assessment is consistent with the observation that the individual clades are morphologically and ecologically distinct.

Not There Yet! Grid-Based Model Predicts 452 Species

Although the number of 126 species in *Cora* (116) and *Corella* (10) now recognized is staggering, it may still be far from the actual species richness in this group. The 376 sequenced samples were gathered in only 209 grids (9%). Only two species-rich grids (1%) could be considered well sampled (Table 1): grid 51 in the Colombian Andes (37 species) and grid 153 in the Atlantic Rain Forest in southeastern Brazil (14 species). These two grids alone account for 51 or nearly half of the species, with 43 species unique to either grid and zero overlap between the two grids. Several grids covering Costa Rica and the Galápagos Islands (41, 42, and 59) could be considered well sampled in terms of available specimens but were set to an intermediate sampling score because only part of the material was recent enough to allow for successful DNA extraction.

Nonlinear estimation between grid species number, T_{grid} , and a combined score of grid biome diversity, B_{grid} , and grid sampling effort, E_{grid} , resulted in a strongly and significantly correlated regression model: $T_{\text{grid}} = 0.52 + e^{(-0.34 + 0.13 \times B_{\text{grid}} \times E_{\text{grid}})}$ ($r = 0.95$, $P = 0.0000$; proportion of variance explained = 91%). We used this model to predict species numbers for each of the 209 grids assuming optimal sampling effort by setting the sampling score, E_{grid} , to the maximum (maximum score = 3) for each grid. This approach resulted in predicted species numbers per grid ranging between 1 and 36 (Table 1), for a total grid sum, G_{total} , of 1,201 species. The highest species number per grid (36) was

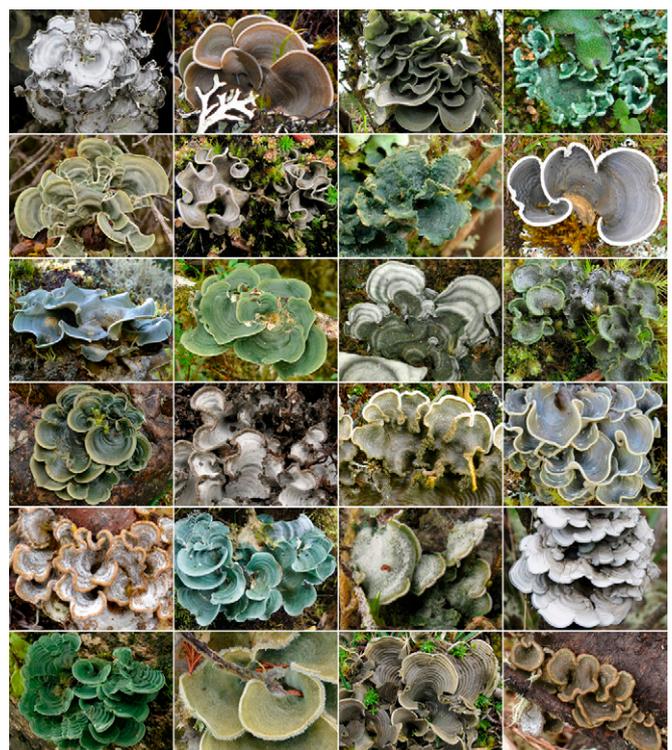


Fig. 2. Field photographs of selected species of *Cora*, showing the remarkable diversity of morphological features. Many of these characters cannot be assessed in herbarium material.



Fig. 3. Grid map of Central and South America and the Caribbean used to predict total species richness of *Cora* and *Corella* species. Sampled grids from which sequences were obtained are marked in white. Black grids are those well-sampled with either high observed species richness (51, 153) or documented absence of species (109). Biogeographical regions with presumed high species richness are indicated. Grid numbers range from 1 to 209 and are given in two-digit numbers; after grid 99, grids 00, 01, etc. mean 100, 101, etc.; after second grid 99, grids 00, 01, etc. mean 200, 201, etc.

predicted for 12 grids and the second highest (17) for 13 grids, for a total of 25 high-richness grids. Low-richness grids included 69 grids with one species per grid; zero species were not predicted for any grid, even if one well-sampled grid yielded zero species. High-richness grids were concentrated in the northern and southern tropical Andes, northern Central America, and the southern Atlantic Rainforest; low-richness grids were mostly found in areas with dominance of lowland rain forest, savanna, or semiarid vegetation. All high-richness grids included either paramo or (mixed) montane or cloud forest vegetation.

Total species number for the target area, T_{total} , is a function of total grid sum, G_{total} , and average distribution of species across grids, $D_{average}$: $T_{total} = G_{total}/D_{average}$. This formula is based on Whittaker's (38) definition of beta diversity or species turnover, originally defined through the following formula: gamma diversity = average alpha diversity \times beta diversity. Beta diversity is then inversely proportional to $D_{average}$ through the following formula: beta diversity = number of grids/ $D_{average}$. Based on the data from the 26 sampled grids (Table 1), observed $D_{average}$ was 1.21 grids per species, which would result in $T_{total} = 1,201/1.21 = 993$ predicted species for the target area. However, because additional data will not only add further species but will increase grid distribution per species, we first used grid interpolation to correct for this bias. For example, if a paramo species was sampled in grids 45, 50, 51, and 136 (four grids), it should also be present in grids with paramo vegetation located in between these grids (additional seven grids). Using this technique, $D_{average}$ increased to 1.42 grids per species, for a predicted total species richness $T_{total} = 1,201/1.42 = 846$ species. We still considered this an overestimate, because interpolation does not account for possible

range extensions of species outside the observed grid area. Therefore, we used an even more conservative approach to estimate $D_{average}$: for the 26 sampled grids, we computed predicted species grid sum G_{26} assuming maximum sampling effort. We then took the observed species number for these grids (126), conservatively assuming that complete sampling would not add any new species, and assumed this to be the same as the total number of predicted species, T_{26} . Average grid distribution per species was then computed as $D_{26} = G_{26}/T_{26} = 335/126 = 2.66$.

Using this value, the prediction for the total number of species for the entire grid map resulted in $T_{total} = 1,201/2.66 = 452$ taxa in *Cora* and *Corella*. This number would be a further remarkable increase from the 126 species recognized in our current phylogeny and an unthinkable dramatic increase from a single species, *D. glabratum*, accepted until a decade ago (11, 12, 16). With more than 400 species, *Cora* would become one of the largest genera of lichenized Fungi. It would also be the first large genus among any group of Fungi almost entirely elucidated "from scratch" using molecular sequence data.

How Realistic Is This Prediction?

Until 10 y ago, the scientific community was unaware that *D. glabratum* represented at least 16 different species in two genera (13, 15). No one could have anticipated that this number would rise to 126 species in our current dataset and even up to 170 species using a coalescent-based species recognition method. Hence, a figure of 452 predicted species does not seem out of the realm of possibility, considering that our current data are still based on very limited sampling effort. The extreme alternative

Table 1. Grids with sequenced specimens of *Cora* and/or *Corella*, with an additional six well-sampled grids in which presence of these genera could not be demonstrated

Grid	Political unit	CBS	SS	CS	SO	SP
11	United States (Florida)	0	2	0	0	1
20	Mexico	2.1	1	2.1	1	2
41	Costa Rica	7	2	14	13	11
42	Costa Rica/Panama	7.7	2	15.4	9	15
45	Colombia/Venezuela	10	2	20	8	36
50	Colombia	7	2	14	9	11
51	Colombia	10	3	30	37	36
52	Colombia	10	2	20	6	36
59	Galápagos Islands	3.5	2	7	2	3
60	Colombia/Ecuador	10	2	20	8	36
70	Ecuador/Peru	10	2	20	16	36
82	Ecuador/Peru	10	1	10	1	36
100	Brazil (Rondonia)	0	2	0	0	1
101	Brazil (Rondonia)	0	2	0	0	1
108	Brazil (Sergipe)	2	1	2	1	2
109	Brazil	0	3	0	0	1
114	Brazil/Bolivia	0	2	0	0	1
115	Brazil (Rondonia)	0	2	0	0	1
124	Peru	8	1	8	4	17
136	Peru/Bolivia	8	2	16	11	17
153	Brazil	8	3	24	14	17
162	Brazil	3.5	2	7	7	3
169	Brazil	6	2	12	4	8
170	Brazil	3.5	1	3.5	4	3
177	Brazil	2.8	1	2.8	1	3
208	Chile/Argentina	0.7	1	0.7	1	1

CBS, corrected biome score (taking into account biome diversity, suitability for occurrence of *Cora* and *Corella*, and substantial presence of ocean surface); CS, combined score; SO, species observed; SP, species predicted; SS, sampling score (for molecular phylogenetic analysis). All other grids were not sampled and had zero species observed and one species predicted per grid.

of species richness and the consideration of lichens due to their ecosystem functions as biofertilizers and water supply regulators (46–48). Preservation of organismic diversity requires not only information about known diversity, but also predictions of unrecognized diversity at a global level. Our approach provides a way of identifying groups that harbor undescribed species and the potential number of species they contain and where to focus future sampling efforts to properly catalog this unrecognized diversity.

Methods

Material and Assessment of Phenotype Characters. ITS fungal barcoding sequences were obtained from a total of 356 new samples of *Cora* and *Corella* collected throughout Central and South America (Table S1). Where possible, samples were documented in the field using high-quality photographs to record potentially important phenotype features. Dried samples were studied in the herbarium using a standardized matrix of 186 characters to assess phenotypic variation, including morphology, internal anatomy, and secondary chemistry (15).

Molecular Sequencing and Phylogenetic Analysis. Genomic DNA was extracted from lichenized thalli, and ITS sequences were generated via Sanger sequencing following previously published protocols (13). Samples that did not yield high-quality Sanger sequences were subjected to 454 pyrosequencing to obtain sequences of the target mycobiont (37). Newly generated sequences were assembled with sequences from GenBank and aligned with MAFFT using the auto option (49) and then manually corrected, resulting in an alignment length of 822 bases. The dataset was subjected to a maximum likelihood (ML) search using RAXML 7.2.6 on the Cipres Science Gateway server (50, 51), with parametric bootstrapping using 500 replicates under the GTRGAMMA model. Both alignment and best-scoring tree file with branch lengths and bootstrap support values were submitted to TreeBASE (accession no. 15783).

Single-Gene Species Recognition. We used GMYC in the *splits* package for R for phylogenetic species recognition, using both the single and multiple threshold approach (29). The ultrametric tree was reconstructed using a relative molecular clock in BEAST 1.7.5 (52), with the following specifications: the general time-reversible substitution model with base frequencies estimated and Gamma and invariant sites with six Gamma categories; speciation through a Yule process with the “yule.birthRate” prior set to an exponential distribution with 4.0 as mean; and the “ucl.d.mean” prior (mean substitution rate) set to an exponential distribution with 0.001 as mean. Estimation of priors was approximated by first running a strict clock and using the “meanRate” posterior estimates as prior for a second run applying a relaxed clock, with all other priors set to default values, and then a third, final run, using the posterior estimates from the second run for “ucl.d.mean” and “yule.birthRate” as priors. All runs were performed with 10 million generations on the Cipres Science Gateway server (51).

Prediction of Total Species Richness. We generated a grid map of the target area, using a grid width of 10° subdivided into three grids each (3.33°), resulting in 209 grids of ~370 km width and 300–370 km height depending on distance to the equator (Fig. 1). For each grid, we determined the number of biomes present.* Each biome received a score approximately proportional to the number of species of *Cora* and *Corella* found in this biome type in our dataset, taking the values of 0 (desert, thornbush, dry forest, lowland grassland and savanna, lowland rain forest), 1 (tundra, temperate forest, cerrado), 2 (temperate rain forest, mixed montane forest including pine forests in Mexico and *Araucaria* forests in southern Brazil, montane rain forest), 3 (cloud forest, puna), and 5 (wet paramo), and for each grid, the

total score was determined by adding individual scores of biomes present. To adjust grids in which more than half of the area was ocean, a correcting factor of 0.7 was applied. For each grid, we set a sampling score, ranging from 0 (not sampled), to 1 (random opportunistic sampling of single sites), 2 (intermediate), and 3 (systematic sampling of more than three sites). The sampling score referred only to material for which we could obtain sequence data and not to the overall sampling effort, because material older than 2 y will usually not yield good DNA extracts. A combined biome-sampling score was then calculated by multiplying the biome score (corrected for ocean-containing grids) with the sampling score (Table S2).

We calculated the number of observed species for each sampled grid, for 26 grids in total (the other 183 grids did not contain sequenced samples). A nonlinear model between species number and biome-sampling score per grid was computed in STATISTICA 6.0, using exponential growth regression with the Quasi-Newton as estimation method, a maximum of 50 iterations, and 0.00099 as the convergence criterion. This model was used to predict species numbers per grid by setting the sampling score to maximum (3). To predict the total number of species, T_{total} , for the entire grid map, we made use of the fact that T_{total} is a function of the sum of species totals for all grids, G_{total} , and the average number of grids $D_{average}$ in which a species was found: $T_{total} = G_{total}/D_{average}$. Although G_{total} is calculated from the number of predicted species per grid, $D_{average}$ can be estimated from the data, as further explained previously.

We reconstructed species-area curves in PC-ORD 6.0 (53) based on randomized subsampling, with Sørensen as a distance measure, to document patterns of species diversity within grids and between adjacent grids belonging to the same ecoregion. For that, we assembled a species (based on the phylogenetic tree) by sampling site (subdivision of each grid) matrix for the grid that contained the greatest number of species (grid 51 in the northern Andes in Colombia) and also compared adjacent grids 50–52 in the northern Andes and grids 153–162 and 169–170 in the Atlantic Rain Forest in southeastern Brazil.

ACKNOWLEDGMENTS. The following colleagues are thanked for providing material or assisting with fieldwork: A. Beck, F. Beilke, W. R. Buck, R. S. Egan, S. Eliasaro, J. Farfan, A. Gerlach, E. Gumboski, J. E. Hernández, L. Herrera, M. A. Herrera-Campos, H. Jonitz, M. Kukwa, M. P. Marcelli, S. A. Martins, E. Navarro, F. Nugra, T. Paredes, C. Plaza, F. Quesada, A. A. Spielmann, A. Suárez, L. Vargas, Z. Vela, and K. Wilk. Ekaphan Kraichak is thanked for advice with using the *splits* package in R and for providing supplemental code. We are indebted to the Galápagos National Park, especially its technical director, Washington Tapia, for support and specimen export permits. Financial support for this study was provided by National Science Foundation Grant DEB-0841405 to George Mason University: “Phylogenetic Diversity of Mycobionts and Photobionts in the Cyanolichen Genus *Dictyonema*, with Emphasis on the Neotropics and the Galapagos Islands” (principal investigator, J.D.L.; co-principal investigators, R.L. and P.M.G.; local coordinator for Galápagos, F.B.). Material was also collected as part of National Science Foundation Grant DEB-0715660 to The Field Museum: “Neotropical Epiphytic Microlichens—An Innovative Inventory of a Highly Diverse yet Little Known Group of Symbiotic Organisms” (principal investigator, R.L.) and DEB-0206125 to The Field Museum: “TICOLICHEN” (principal investigator, R.L.), as well as two lichen courses as part of the Organization for Tropical Studies (OTS) specialty courses syllabus. This publication is Contribution 2093 of the Charles Darwin Foundation for the Galapagos Islands.

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